## SEASONAL VARIATION IN MYCOFLORA OF UNMILLED RICE IN RELATION TO MYCOTOXINS CONTAMINATION

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## ABSTRACT

About 240 samples from different regions of Warangal district of Andhra Pradesh was analysed by employing dilution plate method and seed plating. Most of the samples of unmilled rice were heavily infested. However, fungi associated varied with the condition of the sample and place of collection. In all 30 fungal species be longing 19 genera could be reported in unmilled rice. Species of *Aspergillus, Penicillium* and *Fusarium* were dominant. *Nigrospora oryza, Phoma sorghina* and *Stachybotrys atra* could be recorded in Kharif season samples only, while *Myrothecium roridum* could be recorded in Rabi season samples. In general samples of Kharif season were more mould infested than Rabi season samples. Considerable percentage strains of mycotoxigenic fungi were toxigenic and elaborated aflatoxins, patulin, terreic acid, ochratoxin A, citrinin, zearalenone, DON, roridin, fumonisins and trichothecenes.

Key words: Unmilled rice, Kharif crop, Rabi crop, mycoflora, Aspergillus, Penicillium, Fusarium, mycotoxins.

# INTRODUCTION

Unmilled rice (Oryza sativa L.) is the most important staple food crop in India and the bulk of rice is grown in Kharif or the wet season in Andhra Pradesh. Unmilled rice is the major crop cultivated along the Godavari belt of Andhra Pradesh, India. More relative humidity (%) and warm conditions during rainy season (June to October) favour the mould infestation. Further, seed harvested with high moisture and not dried before storage deteriorates fast due to increased mould activity. The harmful effects of fungal invasion of grain are seed discoloration (Reddy et al., 2005), loss of viability (Reddy et al., 2004; Enikuomehin, 2005) and quality and mycotoxins contamination such as aflatoxins (Liu et al., 2006; Mangala et al., 2006), fumonisins (Silva et al., 2000), trichothecenes (Buck and Cote, 1991), zearalenone and DON (Megalla et al., 2007), citrinin (Nguyen et al., 2007), CPA (Trung et al., 2001), patulin (Rao et al., 2005), ochratoxin A (Makun et al., 2007; Reddy et al., 2007) and Sterigmatocystin (Engelhart et al., 2002). Therefore in the present investigations the seed mycoflora of unmilled rice in relation to season and mycotoxins contamination was studied and discussed in this communication.

#### MATERIALS AND METHODS

An extensive and intensive survey of Warangal district (Fig 1) for fungi associated with unmilled rice during two crop seasons (Kharif and Rabi) in Warangal, Andhra Pradesh (India) was analysed by employing dilution plate method and agar plate method (ISTA, 1993). The fungi thus isolated were subcultured and identified with the help of standard manuals (Samson *et al.*, 1984; Keith, 1996; Singh *et al.*, 1999; Mathur and Kongsdal, 2003; Leslie and Summerell, 2006). The percentage of incidence, frequency and abundance of individual fungus was calculated using the formulae (Ghiasian *et al.*, 2004). The results are obtained are statistically analysed using SPSS software (Version 17.0).

Incidence (%) =	No. of colonies of a species in all the plates			
	Total no. of colonies of all the species in all the plates			
Frequency (%) =	No. of observations in which a species appeared			
requeitey (70) =	Total no. of observation			

Abundance (%) = Total no. of colonies of a species in all observations Total no. of colonies in all observations

Species of *Aspergillus, Penicillium, Fusarium, Myrothecium* and *Trichothecium* which are known to be mycotoxin producers were screened for production of different mycotoxins by as suggested AOAC (1984).

The mycotoxigenic fungi were grown in 25 ml of rice flour medium at  $27\pm2^{\circ}C$  for 15 days. At the end of the incubation period, the culture filtrate was employed for the detection and characterization of different mycotoxins. Liquid-liquid extraction was employed for separation of mycotoxins and different mycotoxins were detected with the help of Thin Layer Chromatography (TLC). On the basis of fluorescence under long wave UV light (360 nm) different mycotoxins were identified. They were further confirmed with help of colour tests and spray reagents as detailed in table 1 (Surekha *et al.*, 2011).

Fig 1: Unmilled rice collected from different pleases of Warangal district



# RESULTS

Variety of moulds were associated with the stored paddy during different times of the year which, however, varied with the age and place of collection of sample (Table 2). During Kharif season unmilled rice seeds supported 30 fungal species representing 19 genera, while during Rabi season it supported 22 fungal species representing 14 genera.

The incidence of species of *Aspergillus, Penicillium* and *Fusarium* occurred through out the year (Fig 2). On the other hand, *A. clavatus, A. flavipes, A.* 

ochraceus, Aureobasidium pullulans, Drechslera spicifer, Paecilomyces lilacinus, Nigrospora oryzae and Phoma sorghina could not be detected during Rabi season. Aspergillus nidulans and Myrothecium roridum were absent in the spermosphere of unmilled rice during Kharif season. The incidence of A. flavus, A. ustus, A. terreus and C. herbarum were recorded during both the crop seasons. On the other hand, A. niger, C. lunata and A. parasiticus could be recorded with comparatively more percentage of incidence during Rabi season, while species of Fusarium were recorded with more percentage of incidence during Kharif season.

A. *flavus* was highest in its percentage of frequency, while *Thielavia terricola* and *Drechslera rostrata* were with least percentage of frequency. A. *terreus, A. niger* and species of *Penicillium* were next highest in their

percentage of frequency. Rest of the fungi occurred with intermediate percentage of frequency. A. flavus was with highest percentage of abundance, while C. herbarum, D. rostrata, N. oryzae and T. terricola were with least percentage of abundance. A. niger, A. terreus, species of Fusarium and Penicillium were next highest in their percentage of abundance.

Name of the	Solvent				
toxin	system	Spray reagent	Detection		
			UV	Visible	
Aflatoxins	C:A (95:5)	-	bl & g	-	
Ochratoxin A	T:Ea:F (6:3:1)	20% AlCl <sub>3</sub>	bb	-	
Patulin	T:Ea:F (6:3:1)	2% phenylhydrazine hydrochloride	-	у	
Terreic acid	T:Ea:F (6:3:1)	Quantitative estimation	-	-	
Deoxynivalenol	C:M (97:3)	P-anisaldehyde, H <sub>2</sub> SO <sub>4</sub> , 20% AlCl <sub>3</sub>	-, ch, bl	у,-,-	
Zearalenone	C:M (97:3)	$Ce(SO_4)_2$ 1% in 6N H <sub>2</sub> SO <sub>4</sub>	-,-,-,br,ch,bl	br,do,lp,-,-,-	
		2,4-DNP, FeCl <sub>3</sub> 3% in ethanol			
		50% $H_2SO_4$ in methanol, $H_2SO_4$ , 20% AlCl <sub>3</sub>			
Roridin E	C:M (97:3)	Phloroglucinal	-	pi	
Citrinin	T:Ea:F (6:3:1)	Ce(SO <sub>4</sub> ) <sub>2</sub> 1% in 6N H <sub>2</sub> SO <sub>4</sub> , 2,4-DNP	у	y,by,lo	
		FeCl <sub>3</sub> 3% in ethanol			
Cyclopiazonic				bl,rb,br	
acid	T:Ea:F (6:3:1)	$Ce(SO_4)_2$ 1% in 6N H <sub>2</sub> SO <sub>4</sub> , 2,4-DNP	У		
		FeCl <sub>3</sub> 3% in ethanol			
		2,4-DNP, FeCl <sub>3</sub> 3% in ethanol, Ammonia			
Ochratoxin A	T:Ea:F (6:3:1)	fumes	-,-,bb	y,pb,-	
			, ,	J 'r - '	
Sterigmatocystin	C:M:A (1:1:1)	20% AlCl <sub>3</sub>	у	-	
Satratoxin H	C:M (97:3)	Phloroglucinal	-	pi	
Trichothecenes	C:M (97:3)	Phloroglucinal	-	pi	

 Table 1: Detection of different mycotoxins in unmilled rice on TLC

**Solvent system:** C=chloroform, A=acetone, M=methanol, T=toluene, Ea=ethyl acetate, F=formic acid. **Detection colours:** g=green, bl=blue, y=yellow, bb=bright blue, ch=charring, pb=purple brown, by=brown yellow, lo=light orange, br=brown, do=dark orange, lp=light purple, pi=pink

#### Statistical analysis: F-Test (ANOVA)

	Kharif season	Rabi season	
Mean	3.41	3.24	
SE	0.942	0.946	
Mean ±SE	3.41±0.942	3.24±0.946	
p-Value	0.43		
Result	Not Significant		

No variation is found in between two crop seasons. Since there is no significance between the mean incidences of two crops and p value is not < 0.05

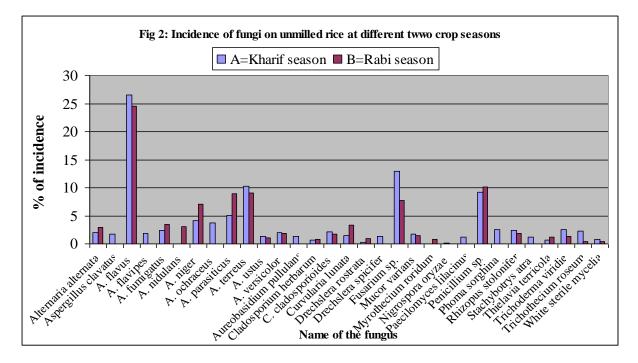
Name of the fungus	Kharif Season			Rabi Season		
	Incidence	Frequency	Abundance	Incidence	Frequency	Abundance
	(%)	(%)	(%)	(%)	(%)	(%)
Alternaria alternata (Fr.) Keissl	1.97	18.3	2.10	2.99	21.1	3.39
Aspergillus clavatus Desm	1.76	14.2	1.47	-	-	-
A. flavus J.H. Friedich Link.	26.6	84.7	20.6	24.5	86.5	26.1
A. flavipes Bain & Sart	1.89	14.2	2.10	-	-	-
A. fumigatus Fresenius	2.36	18.3	3.78	3.41	17.3	2.54
A. nidulans Winter	-	-	-	3.03	19.2	3.39
A. niger Van Tieghem	4.14	40.8	7.78	7.10	30.7	6.36
A. ochraceus Wilhelm	3.77	20.4	3.15	-	-	-
A. parasiticus Spears	5.09	22.4	5.68	8.92	44.2	9.55
A. terreus Thom	10.3	66.5	7.42	9.01	63.5	7.00
A. ustus (Bainier) Thom & Church	1.31	21.7	0.89	1.12	18.0	0.14
A. versicolor (Vuill.)Tiraboschi	1.97	15.8	1.01	1.88	12.6	0.26
Aureobasidium pullulans (De Bary) Arn and herbarum	1.38	14.2	1.10	-	-	-
Cladosporium. herbarum (Pres.) Link.	0.71	1.55	0.11	0.86	1.70	0.19
C. cladosporioides (Fresen.) G.A. de Vries	2.10	16.3	2.31	1.71	7.69	1.69
Curvularia lunata Boedijn	1.43	12.2	1.68	3.36	8.80	4.88
Drechslera rostrata Drechsler	0.23	8.76	0.11	0.98	6.56	0.12
Drechslera spicifer Nelson	1.27	14.2	2.10	-	-	-
<i>Fusarium spp. (F. moniliforme</i> (J. Sheld), <i>F. oxysporum</i> (Schlecht), <i>F. equiseti</i> (Wollenw))	13.0	55.1	10.1	7.71	34.6	8.49
Mucor varians Povah	1.69	14.2	1.47	1.47	11.5	1.27
Myrothecium roridum Tode	-	-	-	0.76	7.69	0.84
Nigrospora oryzae (Berk.& Br.) Petch	0.13	7.12	0.45	-	-	-
Paecilomyces lilacinus Thom	1.24	10.2	1.05	-	-	-
Penicillium spp. (P. citrinum(Thom), P. expansum Link), P. griseofulvum (Dierckx))	9.20	46.9	11.3	10.1	42.3	10.8
Phoma sorghina Breda de Haan	2.52	12.2	1.47	-	-	-
Rhizopus stolonifer (Ehrenb.Fr) Vuill	2.38	22.4	2.94	1.82	30.7	3.82
Stachybotrys atra Corda	1.18	9.61	1.27	-	-	-
Thielavia terricola J.C. Gilman & Abbott	0.64	6.12	0.63	1.24	9.61	1.27
Trichoderma viride Pers	2.48	16.3	2.73	1.36	11.5	1.48
Trichothecium roseum (Pers.) Link.	2.28	16.3	2.73	0.43	3.84	0.42
White sterile mycelia	0.76	6.12	0.89	0.45	6.05	0.69

Table 2: Incidence of different fungi on unmilled rice during two crop seasons (Kharif and Rabi)

Name of the fungus	Kharif Season		Rabi Season		Name of the Toxin	
	<b>S.S.</b>	<b>T.S.</b> (%)	S.S.	<b>T.S.</b> (%)		
Aspergillus flavus	79	30.3	68	29.4	Aflatoxin	
A. ochraceus	23	26.0	-	-	Ochratoxin A	
A. parasiticus	48	27.0	49	28.5	Aflatoxin	
A. terreus	38	44.0	33	33.0	Patulin	
A. terreus	38	31.5	33	3.03	Terreic acid	
A. versicolor	17	23.5	13	15.3	Sterigmatocystin	
Fusarium moniliforme	29	17.4	24	16.6	Zearalenone	
F. oxysporum	31	19.3	22	18.1	Deoxynivalenol	
F. equiseti	27	25.9	17	5.88	Zearalenone	
Myrothecium roridum	18	5.55	15	13.3	Roridin E	
Penicillium citrinum	24	12.5	27	14.8	Citrinin	
P. expansum	25	20.0	24	16.6	Patulin	
P. griseofulvum	27	11.1	23	8.69	CPA	
Stachybotrys atra	29	17.2	-	-	Satratoxin H	
Trichothecium roseum	18	11.1	12	8.33	Trichothecene	

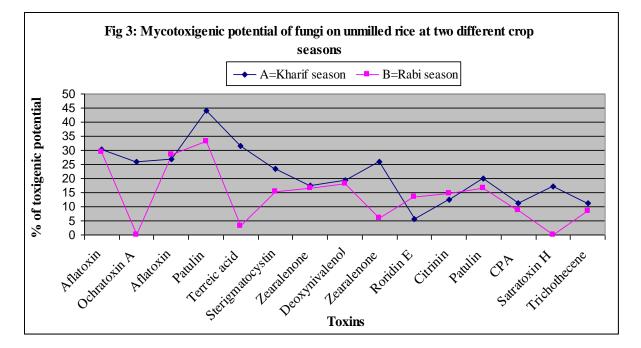
Table 3: Toxigenic potential of unmilled rice in two crop seasons

S.S. = Strains Screened, T.S. = Toxigenic Strains



Rest of the fungi occurred with intermediate percentage of abundance. Almost same trend in the percentage of frequency and abundance was observed during Rabi season. Most of the fungi which are known to be mycotoxigenic elaborated respective mycotoxins (Table 3 and Fig 3). However, percentage of toxigencity varied with the species. About of 30% and 29% of strains of *A. flavus* and *A. parasiticus* respectively elaborated aflatoxins during two crop seasons. *A. ochraceus* which was spotted only during

Kharif season, and above 26% strains elaborated ochratoxin A. Similarly 36% of strains A. *terreus* were toxigenic and produced patulin and terreic acid, while 14% of A. *versicolor* strains were positive for production sterigmatocystin. Strains of different species of *Fusarium* elaborated zearalenone and DON. Similarly about 10% and 17% strains of *M. roridum* and *S. atra* were positive for roridin E and Satratoxin H production respectively. *T. roseum* exhibited its potential to elaborate trichothecene.



About 13%, 18% and 9% strains of *P. citrinum, P. expansum* and *P. griseofulvum* strains were toxigenic and elaborated citrinin, patulin and CPA respectively. From the present investigations it is clear that most of the spermosphere fungi of unmilled rice stored under faulty conditions were toxigenic and could elaborate different mycotoxins. Kharif season was more favorable for mould infestation and mycotoxin contaminations.

# DISCUSSION

The present investigation reveals the wide variations in mycoflora of unmilled rice with place of collection, age of sample and condition of the sample which can be attributed to environmental conditions such as humidity and temperature. Such variation in seed mycoflora has also been reported for different crop seeds studied by them (Fakhrunnisa and Ghaffar, 2006). More mould infestation of unmilled rice collected from flooded area is in agreement with Chary and Reddy (1987) who also recorded many species of *Aspergillus* with comparatively high incidence. Sinha (1987) has also recorded number

aflatoxigenic strains on maize in flooded areas of Bihar. The comparatively more percentage of incidence of moulds as Kharif crop than Rabi crop again may be attributed to warm and humid conditions during rainy season. Sinha (1983) have also recorded more number of fungi on maize crop during Kharif season in Bihar. Comparatively more incidence of fungi during Kharif season may be due humid atmosphere and frequent rain fall. The more incidence of *A. niger, R. stolonifer* 

during Rabi season may be attributed to dry condition prevail during the crop season. No variation is found in between two crop seasons, since there is no significance between the mean incidences of two crops and p value is not <0.05.

**CONCLUSION:** From the present investigations it can be concluded that unmilled rice is prone to moulds infestation which varied with the environmental conditions and condition of stored seeds. The moulds associated with unmilled rice contributed to seed deterioration and constitutional changes. Some of the fungi associated with unmilled rice were mycotoxingenic and potential of elaborating aflatoxins, citrinin, fumonisins, trichothecenes, ochratoxin A, patulin etc.

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#### REFERENCES

- AOAC (Association of Official Analytical Chemists) 1984. Official methods of analysis of the Association of Official Analytical Chemists 14th Edn. Arlington. VA 22209 USA: AOAC.
- Buck, W.B., Cote, L.M. 1991. Trichothecene mycotoxins. In: Hand book of natural toxins, Volume 6. Toxicology of plant and fungal

Marcel Dekker Inc. pp. 523-554.

- Chary, M. P., and Reddy, S. M. 1987. Mycotoxins contamination of dehusked rice in the flooded area of Warangal. Natl. Acad. Sci. Lett. 10(4): 129-132.
- Engelhart S, Loock A, Skutlarek D, et al. 2002. Occurrence of toxigenic Aspergillus versicolor isolates and Maren A Klich 665sterigmatocystin in carpet dust from damp indoor environments. Applied and Environmental Microbiology 68:3886-3890.
- Enikuomehin O. A. 2005. Seed abnormalities and associated mycoflora of rainfed wheat (Triticum aestivum L.) in South Westernn Nigeria. African Journal of Biotechnology Vol. 4 .pp. 672-675.
- Fakhrunnisa, M.H. Hashmi and A. Ghaffar. 2006. Seed-Borne mycoflora of Wheat, Sorghum and Barley. Pak. J. Bot., 38:185-192.
- Ghiasian S.A., Kord-Bacheh P., Rezayat S.M., Maghsood A.H., Taherkhani H. 2004. Mycoflora of Iranian maize harvested in the main production Samson, R.A., Moekstra, E., Van, C.N., 1984. areas in 2000. Mycopathology. 158: 113-121.
- ISTA, 1993. Rules for testing seeds. Seed Sci. Technol., 21: 1-259.
- Keith, S. 1996. Fuskey-Fusarium Interactive Key. Agriculture and Agri-Food Canada, Canada.
- Lislie, J.E., Summerel, B.A. 2006. The Fusarium Laboratory manual. 1<sup>st</sup> ed., Blackwell Publishing Professional, USA, 247 pp.
- Liu, Z., Gao, J., and Yu, J. 2006. Aflatoxins in stored maize and rice grains in liaoning province, china. J. Stored Prod. Res. 42: 468-479.
- Makun, H. A., Gbodi, T. A., Akanya, H. O., Sakalo, A. E., and Ogbadu, H. G. 2007. Fungi and some mycotoxins contaminating rice (Oryza sativa) in Niger state, Nigeria. African J. Biotechnol. 6(2): 99-108.
- Mathur, S.B., Kondgsdal O. 2003. Common laboratory seed health testing methods for detecting fungi. International Seed Testing Association, Switzerland. 234-255.
- Mangala, U. N., Reddy, K. R. N., Singotamu, L., Chary, P. M. S., Reddy, C. S., and Muralidharan, K. 2006. Aspergilli colonize and produce AFB1 in discolored rice grains. J. Mycol. Pl. Pathol. 36(3): 418-426.
- Megalla, S. E., Bennett, G. A., Ellis, J. J., and Shotwell, O. I. 2007. Production of deoxynivalenol and zearalenone by isolates of Fusarium graminearum. Schw. J. Basic Microbiol. 26(7): 415-419.

- compounds. RF Keeler, AT Tu, eds. New York: Nguyen MT, Tozlovanu M, Tran TL, Leszkowicz AP. 2007. Occurrence of aflatoxin B1, citrinin and ochratoxin A in rice in five provinces of the central region of Vietnam. Food Chem. 105:42-47.
  - Rao, G. J., Govindaraju, G., Sivasithamparam, N., and Shanmugasundaram, E. R. B. 2005. Uptake, translocation and persistence of mycotoxins in rice seedlings. Plant Soil. 66(1): 121-123.
  - Reddy, C.S., Reddy, K.R.N., Raja Kumar, N., Laha, G.S., Muralidharan, K. 2004. Exploration of aflatoxin contamination and its management in rice. J Mycol Pl Pathol 34(3):816-820
  - Reddy, K. R. N., Reddy, C. S., and Muralidharan, K. 2007. Exploration of ochratoxin A contamination and its management in rice. Amer. J. Pl. Physiol. 2(3): 206–213.
  - Reddy, K.R.N., Reddy, C.S., Muralidharan, K. 2005. Characterization of AFB1 produced by A. flavus isolated from discolored rice grains. J Mycol Pl Pathol 35(3):470-474.
  - Introduction to food borne fungi. Institute of Royal Netherlands Academy of Arts and Science.
  - da Silva JB, Pozzi CR, Mallozzi MAB. Ortega EM, Correa B. (2000). Mycoflora and occurrence of aflatoxin B1 and fumonisin B1 during storage of Brazilian sorghum. J Agric Food Chem. 48:4352– 4356.
  - Singh, K., Frisrad, J.C., Thrane, U., Mathur, S.B. 1999. An Illustrated manual on identification of someborne Aspergilli, Fusaria, Penicillia and their mycotoxins. Danish Govt. Institute of Seed Pathology for Developing Countries. Denmark. 6-122.
  - Sinha, K.K 1983. Aflatoxin problem in storage and standing maize crop, p. 23-36. In K.S. Bilgrami, T. Prasad and K.K. Sinha (ed.), Mycotoxins in food and feed. Allied Press, Bhagalpur. India.
  - Sinha, K.K 1987. Aflatoxin contamination of Maize in flooded area of Bhagalpur, India. Applied and Environmental Microbiology. 53:1391-1393.
  - Surekha. M, Kiran Saini, V. Krishna Reddy, A. Rajendar Reddy and S.M. Reddy. 2011. Fungal Succession in stored rice (Oryza sativa Linn.) fodder and mycotoxin production. Afrin. J. of Biotech. Vol.10. (4). PP: 550-555.
  - Trung, T. S. Y., Bailly, J. D., Querin, A., Bars, P. L. E., and Guerre, P. 2001. Fungal contamination of rice from South Vietnam, mycotoxinogensis of selected strains and residues in rice. Revue. Med. Vet. 152(7): 555-560.